Res. on Crops **22** (1): 53-59 (2021) DOI: 10.31830/2348-7542.2021.036

With two figures Printed in India

Resistance test of *Echinochloa crus-galli* from West Java toward metsulfuron-methyl and penoxsulam

D. KURNIADIE^{1,*}, K. D. PUTRI², R. WIDIANTO³, Y. SUMEKAR¹ AND U. UMIYATI¹

¹Department of Agronomy Faculty of Agriculture Universitas Padjadjaran, Sumedang, 45363, Indonesia *(email: denny.kurniadie@unpad.ac.id)

(Received: February 20, 2021/Accepted: March 12, 2021)

ABSTRACT

Barnyard grass is acknowledged as the most troublesome weed in rice fields in West Java province of Indonesia due to development of resistance to certain chemical control measures. Therefore, this study was conducted during 2020 at the greenhouse of Faculty of Agriculture, Universitas Padjadjaran, Indonesia aimed to confirm and classify the presence of herbicide-resistant of *Echinochloa crus-galli* toward metsulfuron-methyl and penoxsulam. The resistance level test of *E. crus-galli* was performed using the Whole Plant Pot Test method laid out under Split Plot Design with 3 replications. The main plot comprised of metsulfuron-methyl herbicide: 0, 1, 2, 4, 8, 16 and 32 g a.i./ha, whereas for the dose of penoxsulam herbicide was 0, 2.5, 5, 10, 20, 40 and 80 g a.i./ha. The subplot was the origin of weed: exposed and not exposed to herbicides. The results showed that *E. crus-galli* from East Karawang was included in to low level of resistance to metsulfuron-methyl with a resistance ratio of 5.88. This study has affirmed resistance to metsulfuron-methyl in *E. crus-galli* for the first time. In contrast, no resistance was observed in *E. crus-galli* samples to penoxsulam.

Key Words: Echinochloa crus-galli, metsulfuron-methyl, penoxsulam, weed resistance

INTRODUCTION

Rice (*Oryza sativa* L.) is the main staple food resource in Indonesia (Setyowati *et al.*, 2018). The second-largest rice producer in Indonesia is West Java Province with productivity of 57.54 q/ha (Badan Pusat Statistik, 2019). However, some efforts to maintain and increase rice productivity usually encounter several problems, one among that of which is a competition between rice plants and weeds. Weeds can reduce the rice yield by 21 - 56% in Pakistan (Rabbani *et al.*, 2011) and 11 - 66% in India (Singh and Maiti, 2016; Gharde *et al.*, 2018; Selvaraj and Hussainy, 2020).

Banyard grass [Echinochloa crus-galli (L.) P. Beauv.] is one of the most serious grass weeds in rice production in the world. Antralina (2012) specified that *E. crus-galli* can diminish the rice yield by 57% in Indonesia. The chemical use of herbicides is one way to control weeds in rice cultivation. Beltran *et al.* (2013) have shown that the utilization of herbicides

is 80% more profitable than conventional weeding. However, the extensive use of herbicides will cause weed resistance. Weed resistance occurs frequently over a long period due to the utilize of the same herbicide or herbicide with the same mode of action (Battel, 2018).

Metsulfuron-methyl is a selective, systemic pre and post-emergence herbicides often used in lowland rice cultivation areas to control the growth of several types of weeds (Rao, 2000). Moreover, penoxsulam is also a widely used herbicide for lowland rice around the world because it is selective, effective, and has a broad spectrum (Johnson et al., 2009). Metsulfuronmethyl and penoxsulam have the same mode of action, i.e., inhibiting the acetolactate synthase (ALS) enzyme with the aim that the chains of amino acids valine, leucine and isoleucine are not formed (Su, 2008; Ross and Childs, 2010). Acetolactate synthase inhibitor herbicide (ALS) is the most common herbicide used in lowland rice cultivation in the world (Tranel and Wright, 2002).

²Bachelor student, Faculty of Agriculture, Universitas Padjadjaran, Sumedang 45363, Indonesia. ³Doctoral student, Faculty of Agriculture, Universitas Padjadjaran, Sumedang 45363, Indonesia.

Echinochloa crus-galli biotypes from 11 nations have been reported to be resistant to herbicides that inhibit acetolactate synthase (ALS) (Heap, 2020). In Jiangsu and Anhui Provinces of China, E. crus galli was reported as being resistant to penoxsulam herbicides from the triazolopyrimidine chemical compound. In addition, cross-resistance exists in the biotype of Anhui province against herbicides that inhibit ALS of all chemical compounds, namely: pyrimidinyl-thiobenzoate, imidazolinone, triazolopyrimidine, sulfonyl-carbonyl-triazolinone, and sulfonylurea (Fang et al., 2019).

In Indonesia, *E. crus-galli* is still not officially registered as a resistant weed in paddy fields. A number of populations of *E. crus-galli* have been found to be resistant to ALS-inhibiting herbicides in West Java province, especially metsulfuron-methyl. However, this report has not been supported by an in-depth analysis, thus resistant weeds in West Java are still unclear. Therefore, this experiment was necessary to identify and classify the presence of herbicide-resistance in *E. crus-galli* for commonly used herbicides *viz.*, metsulfuron-methyl and penoxsulam.

MATERIALS AND METHODS

A field experiment was conducted during August to October of 2020 at the greenhouse of Faculty of Agriculture, Universitas Padjadjaran, Indonesia to confirm the presence of herbicide-resistant of *E. crusgalli* toward metsulfuron-methyl and penoxsulam and classify the resistance level of *E. crus-galli* toward metsulfuron-methyl and penoxsulam.

Plant Materials and Growth Conditions

Seeds of suspected resistant *E. crus-galli* populations were collected from rice fields in Rancaekek (6°57′37.6″S 107°45′53.4″E), East Karawang (6°18′22.0″S 107°19′35.4″E), and Majalaya (6°19′39.7″S 107°21′48.0″E) districts. *E. crus-galli* seeds were also obtained from herbicide-untreated paddy fields in Banyusari (6°16′58.0″S 107°34′06.8″E), subsequently referred to as susceptible biotype. Seeds were stored at 4°C for one month to break dormancy (Chen *et al.*, 2016). The Whole Plant Pot Test system (Burgos, 2015) is the method used in

this study. The plants were cultivated in plastic pots (18 cm in diameter) that were filled with rice paddy field soil. All plants were kept in the greenhouse and watered every 3 days.

Dose Response Experiment

Seedlings have been thinned to 10 plants per pot before herbicide treatment. At the 2-3 leaf stage, herbicides were applied using a YOTO 5 litre pressure sprayer with Anvil flood jet green nozzle 1.2 L/min. Metsulfuron-methyl was applied at 0, 1, 2, 4, 8, 16 and 32 g a.i./ha and penoxsulam was applied at 0, 2.5, 5, 10, 20, 40 and 80 g a.i./ha. These rates are equivalent to 0, 0.25 0.5, 1, 2, 4 and 8 times of the recommended field rate of the products. Recommended rates of metsulfuron-methyl and penoxsulam levels are 4 and 10 g a.i./ha, respectively. Dose-response experiments for each herbicide were conducted based on a completely randomized design with three replicates. The plants were cut on the soil surface twenty-one days after treatment (DAT), oven-dried for 48 h at 80°C (Marambe and Amarasinghe, 2002; Valeriy et al., 2020) and shoot dry weights were recorded.

Data Analysis

The whole-plant dose-response data were exposed to ANOVA utilizing SPSS v. 25.0. Non-linear regressions of equation were used to analyze data (Seefeldt *et al.*, 1995):

$$y = C + \frac{D - C}{1 + (x/I_{50})^b}$$

Where, C = Lower limit, D = Upper limit, b = Slope, and I_{50} = A dosage that gives a 50% response.

An analysis of the dose-response curves was performed using Origin Pro 2016 (Originlab Corp, 2016). For each herbicide, Resistance-Susceptible (R/S) ratio was determined as follows: GR_{50} of the resistant (R) population/ GR_{50} of the susceptible (S) population. According to Ahmad-Hamdani *et al.* (2012), the herbicide sensitivity is further divided into four classes: susceptible (R/S< 2), low resistance (R/S = 2-6), moderate resistance (R/S = 6-12) and high resistance (R/S>12).

RESULTS AND DISCUSSION

Sensitivity to ALS Herbicides

There was a significant difference between assay repetitions in the ANOVA results, hence, the outcomes for the repeat assays were summed. The GR50 values for Rancaekek, East Karawang, Majalaya, and susceptible biotypes were 1.36, 7.59, 2.37 and 1.29 g a.i./ha metsulfuron-methyl, respectively (Fig. 1). The GR50 value with

metsulfuron-methyl was 1.90 times higher (7.59 g a.i./ha) than the recommended application dose (4 g a.i./ha) for East Karawang. Based on the interview results, it was found that there were differences in the frequency of herbicide application in one growing season and the length of land use at each location. The rice field from East Karawang was treated with herbicides twice per planting season with land use reaching 10 years; the frequency of herbicide applications at that location is higher than at other locations.

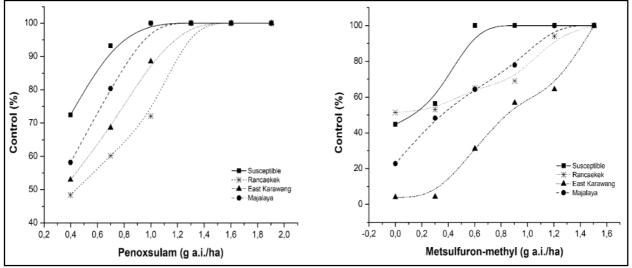


Fig. 1. Inhibition curves by metsulfuron-methyl and penoxsulam from four populations of *E. crus-galli*. Lines are the reaction bends foreseen by non-linear regression; symbols represent shares average survival, upheld the untreated controls.

The GR50 values with penoxsulam were 3.08, 2.46, 2.16 and 1.70 g a.i./ha for Rancaekek, East Karawang, Majalaya, and susceptible biotypes, respectively (Fig. 1). The GR50 values of the E. crus-galli population, which was thought to be resistant were less than the recommended dose of penoxsulam (10 g a.i./ha), which means they were susceptible to penoxsulam. This confirms the information during the interview that the rice fields were treated with metsulfuron-methyl and never with penoxsulam.

Unlike the results of this study, weeds have the potential to develop resistance in the same mode of action against many herbicide families, even though some herbicides have never been used to control weeds. For example, a population of *E. phyllopogon* developed bensulfuron and bispyribac-sodium crossresistance from California. However, only bensulfuron was used in the rice fields and never with bispyribac-sodium (Fischer *et al.*, 2000).

Confirmation of Resistance

The resistance levels of the ALSresistant accessions were calculated as the R/S ratios of the GR50 values (Table 1). Echinochloa crus-galli biotypes from Rancaekek and Majalaya were susceptible to metsulfuronmethyl, yet those from East Karawang showed low resistance to methyl metsulfuron (R/S 5.88), suggesting that it was 5.88 times more resistant to metsulfuron-methyl compared to the susceptible population. To date, 33.33% of the West Java population of *E. crus-galli* has a low resistance (R/S = 2-6) and 66.67% are susceptible to metsulfuron-methyl (R/S = <2). Nevertheless, all biotypes were susceptible to penoxsulam. Based on the GR50 value, penoxsulam R/S ratios for Rancaekek, East Karawang and Majalaya biotypes were 1.81, 1.45 and 1.27, respectively. The East Karawang population was resistant to metsulfuronmethyl but was susceptible (not cross-

Herbicide	Population	\mathbb{R}^2	$\mathrm{GR}_{50}{}^{\mathrm{a}}$	R/S ratiosb
Metsulfuron-methyl	Susceptible	0.87	1.29	_
	Rancaekek	0.81	1.36	1.04
	East Karawang	0.94	7.59	5.88
	Majalaya	0.97	2.37	1.84
Penoxsulam	Susceptible	0.99	1.70	-
	Rancaekek	0.90	3.08	1.81
	East Karawang	0.97	2.46	1.45
	Maialava	0.97	2.16	1.27

Table 1. Metsulfuron-methyl and penoxsulam dose needed to kill 50% of plants (GR50) and R/S ratios obtained from the curves of population S and R E. crus-galli

resistant) to penoxsulam, indicating that penoxsulam may viably be utilized to control this population of *E. crus-galli*.

The different levels of resistance to metsulfuron-methyl and penoxsulam from each location can be seen visually in Fig. 2. Other researchers have identified populations of *Echinochloa crus-galli* resistant to sulfonylurea. *Echinochloa crus-galli* biotype from China is considered to have resistance to flucetosulfuron, propyrisulfuron and rimsulfuron with ratios (R/S) of 16.37, 10.79

and 8.34, respectively (Fang et al., 2019).

Similar to the results of this study, some of the biotypes of *E. crus-galli* (TVA8 and TEV9) from Uruguay were susceptible to penoxsulam with R/S ratios of 1.1 and 1.9 (Marchesi and Saldain, 2019). Furthermore, *E. glabrescens* from the Province of Anhui (China) could be effectively controlled by the oxidant in the field at the recommended dosage level, but not by oxaziclomefone and butachlor. In addition, it was confirmed that the other five species (*E. colonum*, *E. crus-galli* var.

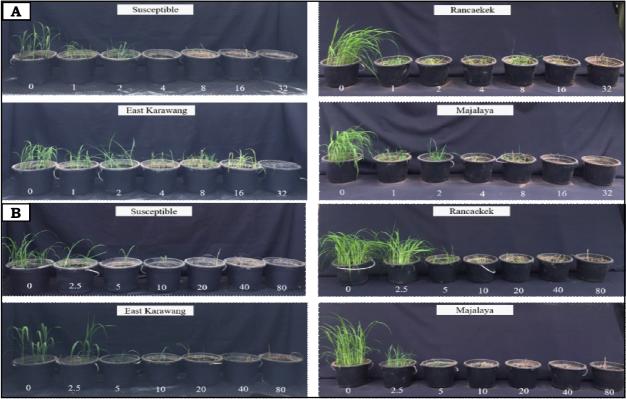


Fig. 2. The growth of resistance (R) and susceptible (S) *E. crus-galli* biotypes at various concentrations of the herbicides metsulfuron-methyl (A) and penoxsulam (B). The dose of each herbicide was in g a.i./ha.

 $^{{}^}aGR_{50}$ alludes to the successful measurements of herbicide causing 50% restraint of dry weight and is demonstrated as grams of active ingredient per hectare (g a.i./ha)

 $^{^{\}mathrm{b}}\mathrm{R/S}$ ratio, resistance index calculated as the ratio of GR_{50} R over GR_{50} S

zelayensis, *E. caudata*, *E. phyllopogon* and *E. oryzoides*) were susceptible to penoxsulam (Wang *et al.*, 2015).

In contrast to the results of this study, a low level of resistance to penoxulam was observed in *E. phyllopogon* (R/S 5) and a nontarget location component was recognized (Yasuor *et al.*, 2009). The lower than anticipated R/S ratio (based on *in vitro*) bioassays, especially for penoxsulam might be due to the polyploidy of *E. crus-galli* and the existence of un-mutated alleles (Panozzo *et al.*, 2013). In addition, *E. crus-galli* biotypes from a few countries have experienced high resistance to penoxsulam with ratios (R/S) of 39 (Turkey) and 69 (China) (Chen *et al.*, 2016; Kacan *et al.*, 2020).

Herbicide resistance mechanisms can be divided into two categories: the target-site resistance mechanism (TSR) and the nontarget site resistance mechanism (NTSR). The mechanism of NTSR includes decreased assimilation or translocation and expanded sequestration or metabolic debasement. On the other hand, TSR mechanisms are advanced processes, unique to a single site of action. The TSR may be attributed to the improved expression of the content of the target site, producing more protein than can be considerably restrained by ordinary application rates of herbicides. Increased expression of the gene can be due to regulatory changes and/or an expanded amount of genomic copy of the target-site gene, which also results in increased transcription (Gaines et al., 2020). In expansion, the results of other studies suggest that mutations in Pro-197 lead to resistance to SU (sulfonylurea) (Tranel et al., 2020). Up to the present, TSR mutation detection has progressively highlighted its significance compared to NTSR although the mechanisms of TSR and NTSR can coexist in the same population (Bai et al., 2019; Chen et al., 2020). For instance, two CYP450 genes, namely CYP94A1 and CYP71A4 constitutively expressed by the short foxtail resistance mechanism to metsulfuron-methyl (Zhao et al., 2019).

Management Implications

In general, the strategies include a mix of early preventive steps and one or more essential control measurements (chemical,

cultivation innovation, biological, and physical). Early preventive steps include identification of resistance and risk evaluation (Liu et al., 2020). In postponing resistance, herbicide rotation has been connected to combat weed resistance (Marochi et al., 2018). For cultivation management, crop rotation is as a rule received by rotating of a grass crop (e.g. rice) with a broadleaf, or by rotating of a wetland crop (e.g. rice) with a dry land crop (Le et al., 2018; Liu et al., 2018). Additionally, logical information shown in bioorganic fertilizers can effectively regulate E. crus-galli and M. vaginalis in rice areas with a normal rate of more than 80% weed concealment (Li et al., 2018). In expansion, the stale seedbed physical action is valuable for depleting the seed bank before planting crops (Feng et al., 2015). The effectiveness of multiple resistant control on E. crus-galliweeds will surpass 85% by using an integrated method (Ma et al., 2012; Zhang et al., 2018).

CONCLUSION

The population of *Echinochloa crus-galli* originating from East Karawang was classified as low resistance to methyl metsulfuron (R/S 5.88) but susceptible to penoxsulam. This study confirms the first case of metsulfuron-methyl (sulfonylurea) resistance in *E. crus-galli*. Meanwhile, the population of *E. crus-galli* originating from Rancaekek and Majalaya was classified as being susceptible to both metsulfuron-methyl and penoxsulam (R/S<2).

ACKNOWLEDGEMNTS

We thank to the Rector Universitas Padjadjaran for the scheme of the Academic Leadership Grant (ALG) for supporting the project.

REFERENCES

Ahmad-Hamdani, M. S., Owen, M. J., Qin Yu and Powles, S. B (2012). ACCase-inhibiting herbicide-resistant *Avena* spp. populations from the Western Australian grain belt. *Weed Technol.* **26**: 130-36.

Antralina, M. (2012). Characteristics of weeds and product components of paddy rice (*Oryza sativa* 1.) SRI system at different weeds presence. *J. Agribisnis dan Pengembangan Wilayah.* **3**: 9-16.

- Badan Pusat Statistik (2019). Harvested area, productivity, and production of rice plants. Indonesia: BPS; [cited 2020 Feb 28]. Available from: https://www.bps.go.id/dynamictable/2019/04/15/1608/luaspanen-produksi-danproduktivitas-padimenurut-provinsi-2018.html
- Bai, S., Zhang, F., Li, Z., Wang, H., Wang, Q., Wang, J. and Bai, L. (2019). Target-site and non-target-site-based resistance to tribenuron-methyl in multiply-resistant *Myosoton aquaticum* L. *Pesticide Biochem. Physiol.* **155**: 8-14.
- Battel, R. (2018). How does herbicide resistance occur? US: MSU Extension Agriculture [cited 2020 Jun. 30]. Available from: https://www.canr.msu.edu/news/how-does-herbicide-resistance-occur
- Beltran, J. C., White, B., Burton, M., Doole, G. J. and Pannell, D. J. (2013). Determinants of herbicide use in rice production in the Philippines. *Agric. Econ.* **44**: 45-55.
- Burgos, N. R. (2015). Whole-plant and seed bioassays for resistance confirmation. *Weed Sci.* **63**: 152-65.
- Chen, G., Wang, Q., Yao, Z., Zhu, L. and Dong, L. (2016). Penoxsulam-resistant barnyard grass (*Echinochloa crus-galli*) in rice fields in China. Weed Biol. Manage. **16**: 16-23.
- Chen, J., Chu, Z., Han, H., Goggin, D. E., Yu, Q., Sayer, C. and Powles, S. B. (2020). A Val-202-phe α-tubulin mutation and enhanced metabolism confer dinitroaniline resistance in a single Lolium rigidum population. Pest Manage. Sci. 76: 645-52.
- Fang, J., Liu, T., Zhang, Y., Li, J. and Dong, L. (2019). Target site-based penoxsulam resistance in barnyard grass (*Echinochloa crus-galli*) from China. *Weed sci.* **67**: 281-87.
- Feng, L., Chen, G. Q., Tian, X. S., Yang, H. M., Yue, M. F. and Yang, C. H. (2015). The hotter the weather, the greater the infestation of *Portulaca oleracea*: opportunistic life-history traits in a serious weed. *Weed Res.* **55**: 396-05.
- Fischer, A. J., Ateh, C. M., Bayer, D. E. and Hill, J. E. (2000). Herbicide-resistant *E. oryzoides* and *E. phyllopogon* in California *Oryza sativa* Fields. *Weed Sci.* **48**: 225-30.
- Gaines, T. A., Duke, S. O., Morran, S., Rigon, C. A., Tranel, P. J., Küpper, A. and Dayan, F. E. (2020). Mechanisms of evolved herbicide resistance. J. Biol. Chem. 295:10307-0330.
- Gharde, Y., Singh, P. K., Dubey, R. P. and Gupta, P. K. (2018). Assessment of yield and economic losses in agriculture due to weeds in India. *Crop Protection* **107**: 12-18.
- Heap, I. (2020). The International Survey of

- Herbicide Resistant Weeds. US: Weed Science; [cited 2020 Jun 30]. Available from: www.weedscience.org.
- Johnson, T. C., Martin, T. P., Mann, R. K. and Pobanz, M. A. (2009). Penoxsulam structure activity relationships of triazolopyrimidine sulfonamides. *Bioorganic Med. Chem.* 17: 4230-240.
- Kacan, K., Tursun, N., Ullah, H. and Datta, A. (2020). Barnyard grass (*Echinochloa crusgalli* (L.) P. Beauv.) resistance to acetolactate synthase-inhibiting and other herbicides in rice in Turkey. *Plant, Soil Environ.* **66**: 357-65.
- Le, D., Nguyen, C. M., Kumar, B. V. N. and Mann, R. K. (2018). Weed management practices to control herbicide-resistant *Echinochloa crus-galli* in rice in Mekong Delta, Vietnam. *Res. Crops* **19** : 20-27.
- Li, Z., Li, D., Zhou, S., Zhou, X. and Bai, L. (2018). The Weeds control of a novel bioorganic fertilizer and its effects on agronomic traits of rice. *Int. J. Agric. Biol.* **20**: 507-12.
- Liu, X., Merchant, A., Xiang, S., Zong, T., Zhou, X. and Bai, L. (2020). Managing herbicide resistance in China. *Weed Sci.* **69**: 1-45.
- Liu, X., Zong, T., Li, Y., Zhou, X. and Bai, L. (2018). Effect of environmental factors on seed germination and early seedling emergence of Carolina geranium (Geranium carolinianum). Planta Daninha 36: doi.org/10.1590/s0100-83582018360100136.
- Ma, G., Bai, L., Liu, D., Liu, X., Lu, Y. and Yu, L. (2012). Control effects of different herbicide combinations on the resistance weeds in direct seeding rice fields. *Chin. Agric. Sci. Bull.* **28**: 195-99.
- Marambe, B. and Amarasinghe, L. (2002). Propanil-resistant Barnyardgrass [Echinochloa crus-Galli (L.) Beauv.] In Sri Lanka: seedling growth under different temperatures and control. Weed Biol. Manage. 2: 194-99.
- Marchesi, C. and Saldain, N. E. (2019). First report of herbicide-resistant *Echinochloa crus-galli* in Uruguayan rice fields. *Agron.* **9** : doi.org/10.3390/agronomy9120790.
- Marochi, A., Ferreira, A., Takano, H. K., Oliveira Junior, R. S. and Ovejero, R. F. L. (2018). Managing glyphosate-resistant weeds with cover crop associated with herbicide rotation and mixture. Ciencia e Agrotecnologia 42: 381-94.
- Originlab Corp. (2016). Origin Graphing & Analysis. US: OriginLab [cited 2020 Jun 30]. Available from: https://www.originlab.com/2016.
- Panozzo, S., Scarabel, L., Tranel, P. J. and Sattin, M. (2013). Target-site resistance to ALS inhibitors in the polyploid species *Echinochloa crus-Galli. Pesticide Biochem. Physiol.* **105**: 93-101.

- Rabbani, N., Bajwa, R. and Javaid, A. (2011). Interference of five problematic weed species with rice growth and yield. *Afr. J. Biotechnol.* **10**: 1854-862.
- Rao, V. S. (2000). Principles of weed science. Science publishers, Inc., New Hampshire, USA.
- Ross, M. A. and Childs, D. J. (2010). Herbicide Mode of Action. US: Department of Botany and Plant Pathology, Purdue University; [cited 2020 Feb 28). Available from: www.bio5.rwth-aachen.de.
- Seefeldt, S. S., Jensen, J. E. and Fuerst, E. P. (1995). Log-logistic analysis of herbicide dose-response relationships. *Weed Technol.* **9**: 218-227.
- Selvaraj, A. and Hussainy, S. A. H. (2020). Evaluating weed management practices for direct sown drum seeded rice (*Oryza sativa*) : A review. *Crop Res.* **55**: 139-51.
- Setyowati, M., Irawan, J. and Marlina, L. (2018). Agronomic characteristics of some Aceh local rice. *J. Agr. Lestari.* **4**: 36-50.
- Singh, V. P. and Maiti, R. K. (2016). A review on factors affecting crop growth in rice (*Oryza sativa* L.). **1**: 101-14.
- Su, S. Q. (2008). New generation herbicide for rice fields-penoxsulam. *World Pesticides* **30**: 48-49.
- Tranel, P. J. and Wright, T. R. (2002). Resistance of weeds to ALS-inhibiting herbicide: What we have learned. *Weed Sci.* **50** : 700-12.
- Tranel, P. J., Wright, T. R. and Heap, I. M. (2020). Mutations in herbicide-resistant weeds to

- ALS inhibitors. US: Weed Science [cited 2020 Jun 30]. Available from: www.weedscience.com.
- Valeriy, A. B., Alexander, F. P., Ekaterina, S. B., Ameth, D., Aleksandr, S. B., Meisam, Z. and Vadim, G. P. (2020). Efficiency of advanced sprayers for nutrient and pesticide application under precision cultivation of spring rapeseed (*Brassica napus*). Res. on Crops 21: 466-72.
- Wang, Q., Chen, G., Jiang, Y., Wang, Q., Yao, Z. and Dong, L. (2015). Sensitivity of *Echinochloa* species to frequently used herbicides in paddy rice field. *J. Nanjing Agric. Univ.* 38: 804-09.
- Yasuor, H., Osuna, M. D., Ortiz, A., Saldain, N. E., Eckert, J. W. and Fischer, A. J. (2009). Mechanism of resistance to penoxsulam in late water grass [Echinochloa phyllopogon (Stapf) Koss.]. J. Agric. Food Chem. 57: 3653-660.
- Zhang, J., Ma, G., Zhou, Z., Tang, W., Yu, X. and Lu, Y. (2018). Application of mechanical weeding technology of direct seeding rice with "seeding and spraying synchronization" in early rice production in Hunan. *Agric. Sci.* **4**: 56-59.
- Zhao, N., Yan, Y., Ge, L. A., Zhu, B., Liu, W. and Wang, J. (2019). Target site mutations and cytochrome p450s confer resistance to fenoxaprop-p-ethyl and mesosulfuronmethyl in *Alopecurus aequalis*. *Pest Manage*. *Sci.* **75**: 204-14.